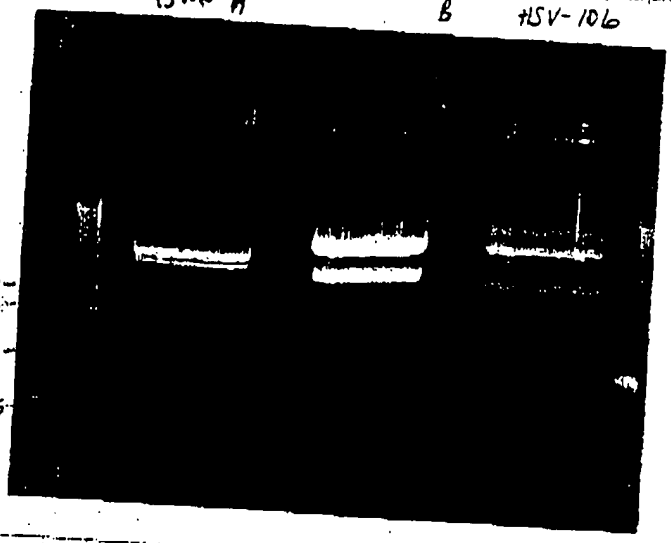


A 15.90 A: BspHI never cut
even w/ o/n & extra
enzyme = β-gal - Xb
took upper band

p 15.90 B XbaI complete en
took upper band

pHSV106 p15A1
took band at ~ 2.2 kb

did Glassmilk, NaI
counted. Eluted w/ 20µl x
H2O and freez = o/n

5µl of each sample for concentration est

1. 15.90 A = 100ng/5µl = ~~20ng~~ 20 ng/µl = β-gal
want ~~50ng~~ 34.2 ng for ligation ≈ 1.5 µl

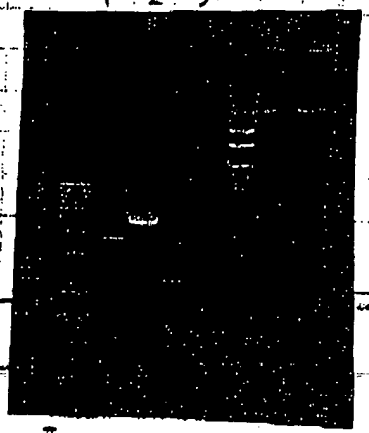
2. 15.90 B = XbaI/HindIII P3E
≈ 0.6µg/5µl = ~~100ng~~ 120 ng/µl
want 45 ng ≈ 0.5 µl

3. p15A1 = HSV-tk gene
≈ 10 ng/µl want 20 ng ≈ 2 µl

Vector ~~Eco~~ 15.81A use 2µl of ~~1:5~~ 1:5 dil

Vector 15.81B use 2µl of 1:5 dil

pcDNA p16.7L want 18 ng use 2µl of 1:100



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ligate a) vector p15.81b — β-gal into BS as XbaI
insert p15.90a

b) vector p15.81a — pSO to BS as XbaI (blunt)/HindIII
insert p15.90b — into HincII/HindIII BS [illegible]

c) ~~vector~~ insert p15.91 — HSV-tk into pcDNA3 as BglII/EcoRI
~~insert~~ vector p16.71 — into BamHI/EcoRI

Cecil set-up the ligation last evening @ 14°C o/n
this morning - run ligation control gel
- plate on 50 ng/ml Amp° run 1, 2, 5-16

yesterday I also retransfected the four additional clones from kit Microbix, Toronto, CN. They failed to grow again ... not much there this morning. I will try one last time today using the ligation experiment as a control.

Also want to characterize the following plasmids from kit that came out of CsCl yesterday

pΔE1sp1B (lower band from CsCl)
use 2 µl/digest
pXC1 (lower band CsCl)
use 2 µl/digest
pBHG 10 or 11 ← this doesn't look as good as others
use 5 µl/digest

note: pΔE1sp1a doesn't look too good at all & needs to be regrown

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